Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

4TH EDITION



Gregory J. Bugbee, Summer E. Stebbins, Martha E. Barton, and Jordan A.G. Wostbrock

The Connecticut Agricultural Experiment Station Department of Environmental Science & Forestry



123 Huntington Street New Haven, CT 06511





The Connecticut Agricultural Experiment Station was founded in 1875. It is chartered by the General Assembly to make scientific inquiries and conduct experiments regarding plants and their pests, insects, soil to make and water, and to perform analyses for state agencies. Station laboratories or research farms are located in New Haven, Windsor, Hamden, and Griswold.



Equal employment opportunity means employment of people without consideration of age, ancestry, color, criminal record (in state employment and licensing), gender identity or expression, genetic information, intellectual disability, learning disability, marital status, mental disability (past or present), national origin, physical disability (including blindness), race, religious creed, retaliation for previously opposed discrimination or coercion, sex (pregnancy or sexual harassment), sexual orientation, veteran status, and workplace hazards to reproductive systems unless the provisions of sec. 46a-80(b) or 46a-81(b) of the Connecticut General Statutes are controlling or there are bona fide occupational qualifica-tions excluding persons in one of the above protected classes. To file a complaint of discrimination, con-tact Dr. Jason White, Director, The Connecticut Agricultural Experiment Station, 123 Huntington Street, New Haven, CT 06511, (203) 974-8440 (voice), or Jason.White@ct.gov (e-mail). CAES is an affirmative action/equal opportunity provider and employer. Persons with disabilities who require alternate means of communication of program information should contact the Chief of Services, Michael Last at (203) 974-8442 (voice), (203) 974-8502 (FAX), or Michael.Last@ct.gov (e-mail).

Acknowledgments

The efforts of Dr. Robert Capers, Ms. Nancy Murray, Ms. Roslyn Reeps, Ms. Amy Weiss, Ms. Calanthe Cavadini, and Ms. Jennifer Fanzutti are gratefully acknowledged.

Funding provided by the Grants for the Control of Aquatic Invasive Species Program administered by the Connecticut Department of Energy and Environmental Protection (DEEP) as well as the Connecticut Federation of Lakes.

© 2022 The Connecticut Agricultural Experiment Station

Bulletin No. 1087

Table of Contents

Introduction	
How to Use This Guide	
Table of Connecticut's Invasive or Potentially Invasive Aquatic Plants	5
What to Do if You Find a Species Discussed in This Guide	5
State Map of Locations of Invasive Aquatic Plants	6
Additional Plant Identification Resources	6
Plant Terms	7
Plant Species Descriptions:	
Butomus umbellatus, Flowering rush	
Cabomba caroliniana, Fanwort	9
Callitriche stagnalis, Pond water-starwort	
Egeria densa, Brazilian waterweed	11
Eichhornia crassipes, Common water-hyacinth	
Hydrilla verticillata, Hydrilla	13
Iris pseudacorus, Yellow iris	16
Lythrum salicaria, Purple loosestrife	
Marsilea quadrifolia, European waterclover	
Myosotis scorpioides, Forget-me-not	
Myriophyllum aquaticum, Parrotfeather	20
Myriophyllum heterophyllum, Variable-leaf watermilfoil	21
Myriophyllum spicatum, Eurasian watermilfoil	
Najas minor, Minor naiad	
Nelumbo lutea, American water lotus	24
Nymphoides peltata, Yellow floating heart	25
Pistia stratiotes, Water lettuce	
Potamogeton crispus, Curly leaf pondweed	27
Rorippa microphylla, Onerow yellowcress	
Rorippa nasturtium-aquaticum, Watercress	
Salvinia molesta, Giant salvinia	
Trapa natans, Water chestnut	31
Commonly Confused Aquatic Plants	
Invasive Aquatic Plant Identification Key	35
Introduction and Dispersal of Aquatic Invasives	
Managing Nuisance Aquatic Vegetation in Connecticut	
Nuisance & Purportedly Invasive Algae in Connecticut Waterways	43
Invasive Aquatic Clams and Mussels	45
Clam and Mussel Species Descriptions:	
Corbicula fluminea, Asian clam	46
Dreissena bugensis, Quagga mussel	47
Dreissena polymorpha, Zebra mussel	
Commonly Confused Aquatic Mussels	
Literature Cited	

Introduction to Aquatic Plants

Aquatic plants are essential components of healthy ecosystems in lakes and ponds. They cleanse water and provide habitat for beneficial aquatic organisms. Because invasive species are not native, they have few natural enemies. Their dramatic growth rates can degrade native ecosystems, decrease recreational opportunities, and reduce local real estate values (Connecticut Aquatic Nuisance Species Working Group 2006, Fishman et al. 1998, Les and Mehrhoff 1999). Recent vegetation surveys of 256 lakes and ponds by the Connecticut Agricultural Experiment Station Invasive Aquatic Plant Program (CAES IAPP) found invasive plants in nearly 60 percent of the waterbodies (CAES IAPP 2022).

Approximately three-quarters of the invasive aquatic plant species in southern New England were introduced as cultivated plants (Les and Mehrhoff 1999). These introductions come from dumping of aquariums and water gardening. Further spread is caused by recreational boating and plant fragments mixed with live bait used by fishermen (Couch and Nelson 1985). Spread of invasive plants from one lake to another also occurs naturally by wildlife and downstream flow. Once established, eradication of invasive aquatic plants is extremely difficult. Preventing introductions by inspections, public education, early detection, and rapid response is critically important.

This guide is intended to provide information on the identification and distribution of the 22 aquatic plants listed as invasive or potentially invasive (Table 1) by the Connecticut General Statute (Sec. 22a-381d). The Connecticut Invasive Plants Council has identified which invasive species in the General Statute are considered aquatic plants. The sale of these plants, with the exception of common water-hyacinth (*Eichhornia crassipes*) and water lettuce (*Pistia stratiotes*), is also banned by State Statute, and their transport is limited to activities associated with control and education. Fines of up to one hundred dollars can be imposed for each violation. Also included in this edition is information on the identification and distribution of the invasive algae Didymo and invasive clams and mussels.

How to Use This Guide

Identifying Connecticut's freshwater aquatic plants is challenging. CAES IAPP surveys have found over 100 native species and 13 invasive species (Figure 1). We use many references when plant identification is questionable including Crow and Hellquist (2000) and Fassett (1957), other recognized experts, and molecular identification using DNA sequencing. Some of the potentially invasive plants discussed here have never been documented in Connecticut and may be unfamiliar to readers. Certain invasive aquatic plants can be easily confused with native or other invasive plants so care must be taken to ensure accuracy. The places where plants are found are often related to their means of dispersal (Table 1), and sometimes this gives a clue to their identification.

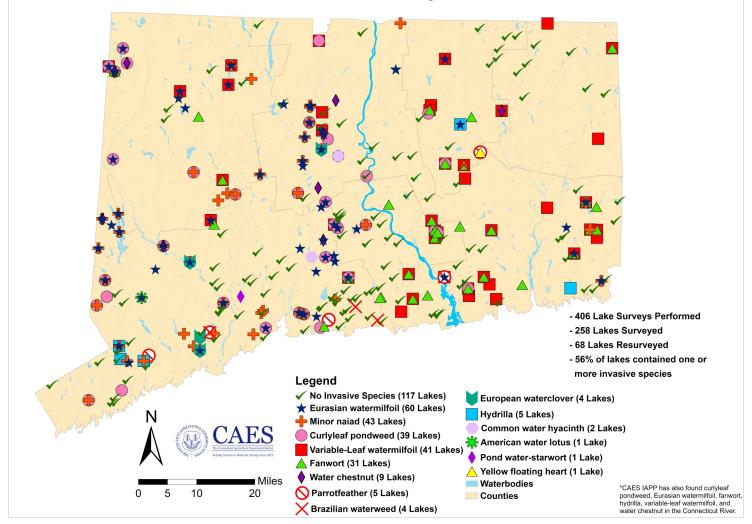
This guide has many parts. Each plant has a summary page containing pictures, a list of key features, and a map of where the plant has been found by either CAES IAPP or the Invasive Plant Atlas of New England (IPANE, 2022). Other sources may have found some of the plants elsewhere, and the maps are not meant to suggest the plants are limited to the locations shown. There is a series of comparative pictures that help differentiate the invasive species from similar native plants, and there is a plant identification key that provides a step-by-step method for narrowing plants to their species. This key also includes native plants that are commonly mistaken for invasive species. A section is included on aquatic plant prevention and control. New to this edition is a section on invasive aquatic clams and mussels.

Table 1. Invasive and potentially invasive aquatic plants listed in the Connecticut General Statutes(Sec. 22a-381d).

. #	Scientific Name	Common Name	Dispersal
1	Butomus umbellatus	Flowering rush	Water Gardening
2	Cabomba caroliniana	Fanwort	Aquariums, Boats/Trailers, Bait
3	Callitriche stagnalis	Pond water-starwort	Water Gardening
4	Egeria densa	Brazilian water-weed, Anacharis, Egeria	Aquariums, Boats/Trailers, Bait
5	Eichhornia crassipes*	Common water hyacinth	Water Gardening
6	Hydrilla verticillata	Hydrilla	Aquariums, Boats/Trailers, Bait
7	Iris pseudacorus	Yellow iris, Yellow flag iris	Nursery Stock, Water Gardening
8	Lythrum salicaria	Purple loosestrife	Nursery Stock, Water Gardening
9	Marsilea quadrifolia	European waterclover, Water shamrock	Water Gardening, Boats/Trailers
10	Myosotis scorpioides	Forget-me-not, Water scorpion-grass	Water Gardening
11	Myriophyllum aquaticum	Parrotfeather	Aquariums, Boats/Trailers
12	Myriophyllum heterophyllum	Variable-leaf watermilfoil	Aquariums, Boats/Trailers
13	Myriophyllum spicatum	Eurasian watermilfoil	Aquariums, Boats/Trailers
14	Najas minor	Brittle water-nymph, Minor naiad	Boats/Trailers
15	Nelumbo lutea	American water lotus	Water Gardening
16	Nymphoides peltata	Yellow floating heart	Water Gardening
17	Pistia stratiotes*	Water lettuce, Tropical duckweed	Water Gardening
18	Potamogeton crispus	Curlyleaf pondweed, Crispy-leaved pond- weed	Boats/Trailers
19	Rorippa microphylla	Onerow yellowcress	Water Gardening
20	Rorippa nasturtium- aquaticum	Watercress	Water Gardening
21	Salvinia molesta	Giant salvinia	Water Gardening
22	Trapa natans	Water chestnut	Water Gardening, Boats/Trailers
	*Plants that are not banned		

What to Do if You Find a Species Discussed in This Guide

Early detection and rapid response are important for preventing invasive aquatic species from becoming future problems. Before taking action, however, it is important that the plant be positively identified and the location accurately recorded. Latitude and longitude coordinates taken with a global positioning system (GPS) are best. Plant samples requiring further identification need to be mailed or taken to the CAES IAPP, 123 Huntington Street, New Haven, CT 06511, or another qualified entity such as the Connecticut Department of Energy and Environmental Protection (CT DEEP). You can call CAES IAPP at (203) -8512 with questions.



Locations of Invasive Plants Found by CAES IAPP 2004-2022

Figure 1. Locations of invasive aquatic plants found by CAES IAPP surveys from 2004-2022.

Additional Resources for Plant Identification

- CAES IAPP web page, aquatic plant survey requests, online herbarium, and reprints of this guide https://portal.ct.gov/caes-iapp
- The Invasive Plant Atlas of New England https://www.eddmaps.org/ipane/
- Invasive Plants of the United States http://www.invasiveplantatlas.org/
- University of Florida, Center for Aquatic and Invasive Plants http://plants.ifas.ufl.edu/

USDA National Invasive Species Information Center http://www.invasivespeciesinfo.gov/

Definitions of Plant Terms

Alternate: leaves not directly across from each other on the stem, one per node Compound: leaf containing many individual leaflets Dissected: leaf divided into many narrow segments; appear feathery, branched or forked Entire: leaf not divided and margins not toothed Forked: leaf divided into two or more equal segments Lanceolate: lance-shaped, long, wider in the middle foliage Leaflet: one of many leaf-like structures that make up a leaf Margin: the edge or border of a leaf Opposite: leaves are directly across from each other on the stem, two per node Petiole: leaf stalk; stem-like structure that attaches a leaf to the stem Rhizome: horizontal underground stem often sending out roots and shoots from its nodes Rosette: a dense cluster of leaves that are all at a single height, like petals of a rose Stolon: horizontal above ground stem often sending out roots and shoots at nodes, also termed "runner"

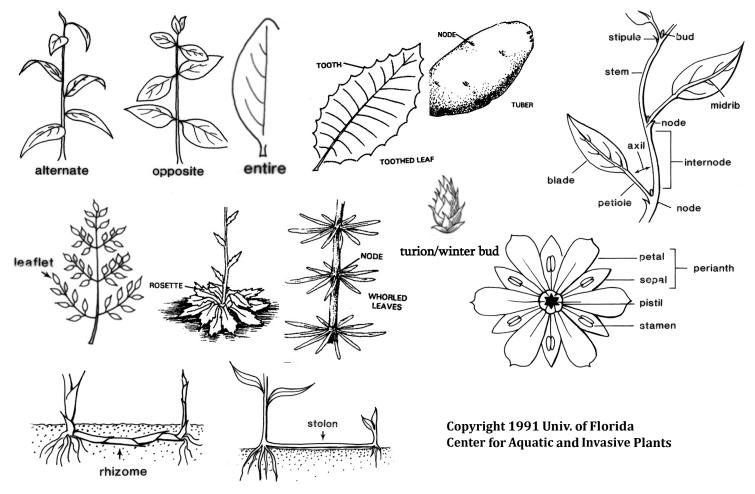
Spike: unbranched continuation of the stem where flowers are located, usually located above the water **Tooth**: points or projections along a leaf margin

Tuber: modified, underground stem for starch storage and a form of vegetative reproduction

Turion: a modified leaf bud on a stem or shoot, a form of vegetative reproduction

Whorled: three or more leaves at the same node, forming a ring-like arrangement

Winter Bud: a modified leaf bud that survives the winter and facilitates vegetative reproduction



Butomus umbellatus

Common name:

Flowering rush

Origin:

East Asia

Key features:

Stems: Can be found along shorelines and into water 9 feet (3 m) deep

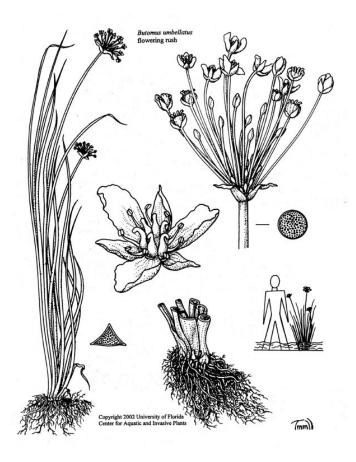
Leaves: Long, narrow, sword shaped leaves up to 3 feet (1 m) tall that originate at base. Leaves are fleshy with twisted ends, grass-like, cross section of leaves are triangular

Flowers: Inflorescence contains pink to white flowers 0.8-1.2 inches (2-3 cm) across with 3 petals and 3 sepals on a stalk that can be 3 feet (1 m) tall

Fruits/Seeds: Fruit is a follicle Reproduction: Seeds and rhizomes

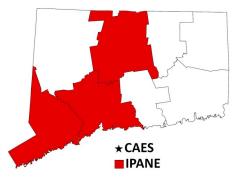
Easily confused species:

Bur-reeds: Sparganium spp.









Page 8 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Cabomba caroliniana

Common names:

Fanwort Carolina fanwort

Origin:

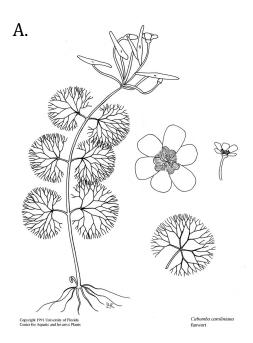
Southeast United States South America

Key features:

Plants are submersedStems: Can be 6 feet (2 m) longLeaves: Dissected, opposite leaves 0.8-2 inches (2-5 cm) are fan-likeand made up of forked leaflets attached to the stem by a petiole.Floating leaves 0.2-0.8 inches (6-20 mm) wide are oblong and pro-duced on flower shootsFlowers: Small, solitary flowers are usually white to pinkishFruits/Seeds: Flask shapedReproduction: Seed and fragmentation

Easily confused species:

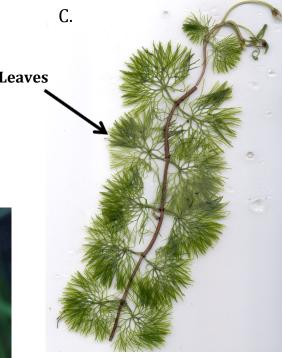
Watermilfoils: *Myriophyllum* spp. White water crowfoot: *Ranunculus longirostris* Water marigold: *Megalodonta beckii*

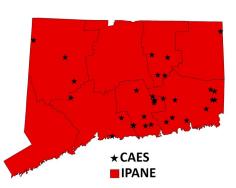




- A. Copyright 1991 Univ. of Florida, Center for Aquatic and Invasive Plants
- B. Copyright 2002 Univ. of Florida, Photo by A. MurrayC. Photo by A. Smagula







Callitriche stagnalis

Common name:

Pond water-starwort

Origin:

Europe North Africa

Key features:

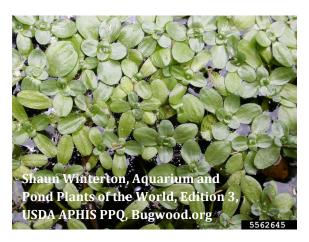
Plants are submersed with floating rosettes **Stems:** 4-12 inches (10-30 cm) long **Leaves:** Floating leaves are opposite and oval or spoon shaped 0.8 × 0.1-0.3 inches (2 cm × 3-8 mm), submerged leaves are narrower and tend to be smaller **Flowers:** Small with 2 small bracts at their base, flowers are close to each other at leaf bases for self-pollination **Fruits/Seeds:** Round 0.06-0.08 inches (1.5-2 mm) thick forming 4 mericarps that have thin winged margins

Reproduction: Cloning and seeds

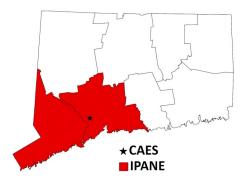
Easily confused species:

Other *Callitriche* spp. (can only distinguish them by their fruit)









Page 10 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Egeria densa

Common names:

Brazilian waterweed Brazilian elodea South American waterweed

Origin:

South America

Key features:

Plants are submersed

Stems: Plant stems green, soft and typically 1-2 ft (0.3-0.6 m) long Leaves: Leaves entire 0.4-1.2 inches (1-3 cm) long by 0.2 in (5 mm) wide, leaves toothed (need magnification), leaves are whorled with typically 4 leaves per whorl Flowers: Small white flowers with three petals, only staminate (male) flowers found in the US Reproduction: Fragmentation

Easily confused species:

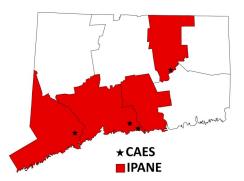
Waterweeds (Native): *Elodea nuttallii* and *E. canadensis* Hydrilla: *Hydrilla verticillata*











Eichhornia crassipes*

Common names:

Common water-hyacinth Floating water-hyacinth

Origin:

Brazil

Key Features:

Stems: Free floating plant

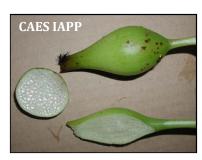
Leaves: Leaves are oval 1.6-4.7 inches (4-12 cm), thick, waxy and form a rosette, petioles are inflated which helps with floatation **Flowers:** Flowers are light purple with one petal having a dark-

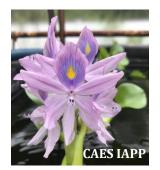
er blotch with a yellow center 2.0-2.8 inches (5-7 cm) **Fruits/Seeds:** Fruit is a capsule with ribbed seeds **Reproduction:** Seeds and stolons

Easily confused species:

None

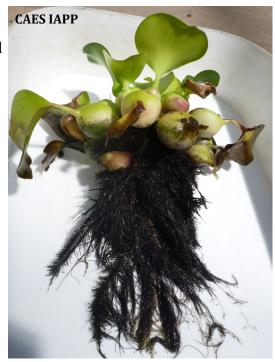
*Not banned for sale in Connecticut

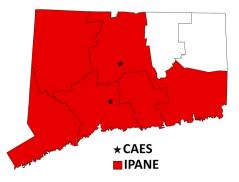












Page 12 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Hydrilla verticillata

Common name:

Hydrilla

Origin:

Asia

Key features:

Plants are submersed

Stems: Slender, branched and up to 25 feet (7.5 m) long **Leaves:** Whorled leaves approx. 0.7 inches (1.5 cm) long, whorls often have 5 leaves (range 4-8); leaf margins are visibly toothed

Flowers: Female flowers have three translucent petals that have reddish streaks; male flowers have three petals and can be white to red in color

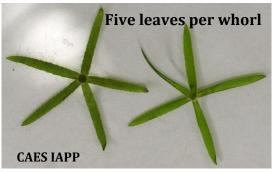
Fruits/Seeds: Small tubers (key feature) can be found in the sediment, turions form along the stem

Reproduction: Fragmentation, turions, tubers and seeds

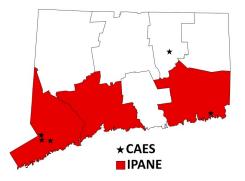
Easily confused species:

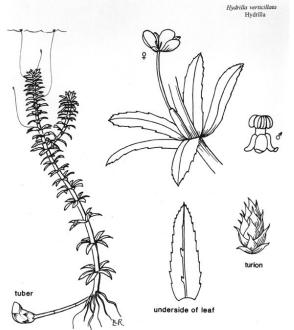
Waterweeds (Native): *Elodea nuttallii* and *Elodea canadensis* Brazilian waterweed: *Egeria densa*











Copyright 1991 Univ. of Florida Center for Aquatic and Invasive^PPlatites Generate.

Hydrilla in the Connecticut River

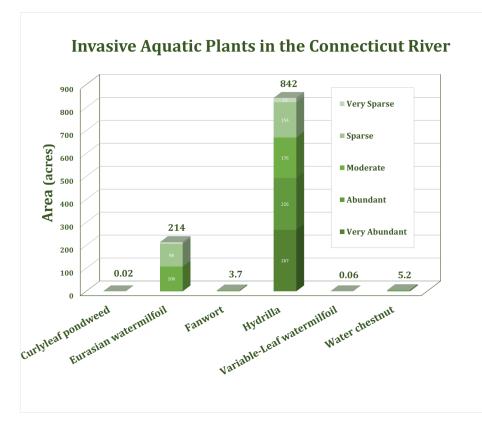
The Connecticut Agricultural Experiment Station Invasive Aquatic Plant Program (CAES IAPP) discovered a new strain of hydrilla the Connecticut River in 2016. Hydrilla is among the most troublesome

invasive aquatic plants in Florida and other southern states. It crowds out native vegetation, harms fisheries, sickens wildfowl, impedes recreation, and reduces property values. CAES IAPP surveys from 2019 - 2022 found 842 acres of hydrilla from Agawam, MA to a mile or so of Long Island Sound. Large dense patches were common in most coves, tributaries, and shallows along the river's mainstem. In some areas, hydrilla spread out over the surface making



Drone image of dense hydrilla in the Mattabesset River in Middletown, CT in 2020. CAES IAPP

navigation nearly impossible. Finding such dense stands of hydrilla in a northern state is alarming. CAES IAPP has found small hydrilla populations in several CT lakes, but these do not compare to the extensive areas



in the Connecticut River. The Connecticut River hydrilla is far more robust than seen elsewhere in the State. This could be a result of river flow, nutrients, or genetics. The Connecticut River hydrilla is a distinct strain from that found anywhere else. Native eel grass beds are extensive throughout the river and create critical habitat for juvenile fish and other aquatic organisms. Hydrilla is encroaching on the eel grass beds with uncertain consequences.

Page 14 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

The Connecticut River hydrilla is genetically distinct from hydrilla elsewhere. Unlike other hydrilla strains it has yet to be observed sprouting from potato-like tubers. Turions, fragments, and possible perennial root systems appear to be its mode of reproduction. This may offer an advantage since tubers can remain in the sediment for years and evade control.

There is great concern that the Connecticut River hydrilla could spread to other bodies of water by boats, canoes, kayaks, and other equipment that can transport fragments of the weed to new locations. Education efforts are underway to assure those using the Connecticut River are aware of the problem and take appropriate "Clean, Drain, and Dry" precautions. In addition, the United States Army Corps of Engineers has recently received funding to conduct hydrilla management demonstration projects with assistance from CAES IAPP and other stakeholders. The hope is a more comprehensive plan for addressing the Connecticut River hydrilla problem.

To Learn More

- Explore a <u>map</u> of the Connecticut River Basin. <u>https://caes.maps.arcgis.com/apps/webappviewer/</u> index.html?id=007f6ee203b74bcbb1d6e68a953d8baf
- <u>Watch a video</u> describing the hydrilla invasion on the Connecticut River. <u>https://</u> <u>vimeo.com/505384824</u>
- Read the Connecticut Agricultural Experiment Station's <u>report</u> on hydrilla and other aquatic plants on the Connecticut River. <u>https://portal.ct.gov/-/media/CAES/DOCUMENTS/Publications/Bulletins/</u> <u>B1084.pdf</u>
- Visit the <u>Connecticut River Conservancy</u> to find out more about hydrilla and the steps each of us can take to help slow its spread. <u>https://www.ctriver.org/get-involved/stopping-an-invasive-species-water-chestnut-2/hyrilla-in-the-ct-river-watershed/</u>

Iris pseudacorus

Common names:

Yellow iris Yellow flag

Origin:

Europe Western Asia Northwest Africa

Key features:

Leaves: Lanceolate leaves are flattened with a raised mid rib and rise out of the soil, parallelvenation, the tips of the leaves are pointed and arch over

Flowers: Flowers are on peduncles 3-4 feet (1-1.3 m) tall. Several light to dark yellow flowers are on each stem with 3 small erect petals and 3 large downward sepals

Fruits/Seeds: Fruit is a capsule, seeds are brown **Reproduction**: Seeds and rhizomes

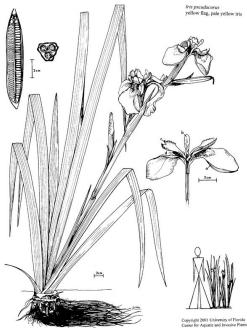
Easily confused species:

Northern blue flag iris: Iris versicolor

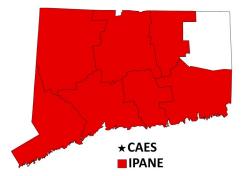


Shaun Winterton, Aquarium and Pond Plants of the World, Edition 3, USDA APHIS PPQ, Bugwood.org





Copyright 2001 Univ. of Florida Center for Aquatic and Invasive Plants



Page 16 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Lythrum salicaria

Common name:

Purple loosestrife

Origin:

Europe

Key features:

Stems: Plants have herbaceous stems and can grow 1.5-5 feet (0.5-1.5 m) tall

Leaves: Opposite, or in whorls of 3, 1-4 inches (3-10 cm) long,

linear, or lanceolate in shape, leaves can be smooth or hairy

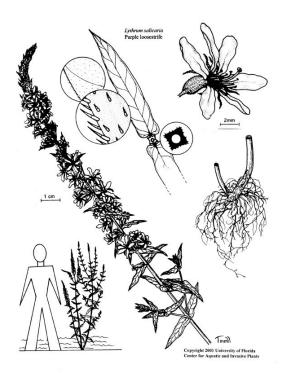
Flowers: Large, pink-purple flowers clustered on long terminal spikes 4-16 inches (10-40 cm) long, floral tube is twice as long as it is wide and typically has 6 petals

Fruits/Seeds: Fruit is a two cavity capsule with numerous reddishbrown seeds

Reproduction: Seed

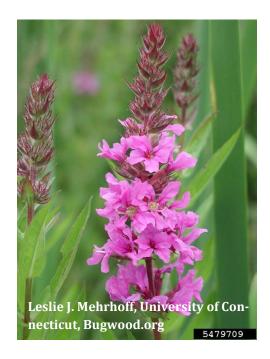
Easily confused species:

Winged loosestrife: Lythrum alatum



Copyright 2001 Univ. of Florida Center for Aquatic and Invasive Plants







Marsilea quadrifolia

Common names:

European waterclover Water shamrock

Origin:

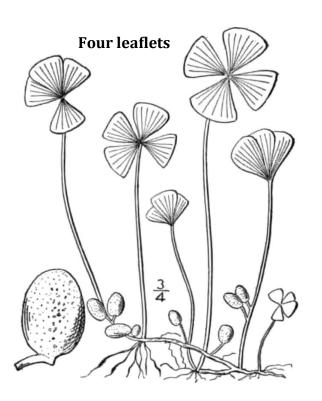
Europe

Key features:

Floating leaf plant **Stems:** Smooth petioles 2-12 inches (5-30 cm) **Leaves:** Comprised of 4 fan-shaped leaflets (similar to a fourleaf clover) **Fruits/Seeds:** 2 or 3 dark brown sporocarps 0.2 inches × 0.2 inches (4-5.5 mm × 3-4 mm) **Reproduction:** Cloning and sporocarps

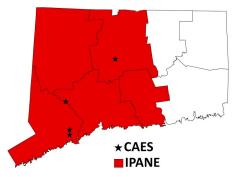
Easily confused species:

None



Britton, N.L., and A. Brown. 1913





Myosotis scorpioides

Common names:

Forget-me-not Yellow eye forget-me-not Water scorpion-grass

Origin:

Europe Western Asia

Key features:

Plants grow 8-24 inches (20-60 cm) in height **Stems:** Stems are angled, often creeping **Leaves:** Lower leaves are tapered to the base while the upper leaves are more oblong, leaves are alternate, with short hairs **Flowers:** Flowers are flat and are typically blue with a yellow center,

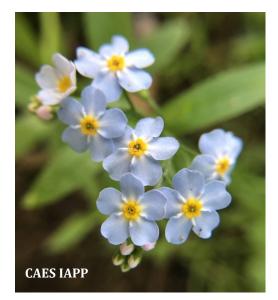
0.2-0.4 inches (6-9 mm) wide, along a simple inflorescence with a common axis

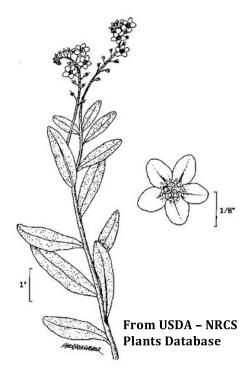
Fruits/Seeds: Seeds are contained in a nutlet that is angled and keeled on the inner side

Reproduction: Seeds

Easily confused species:

Bay forget-me-not: Myosotis laxa









Myriophyllum aquaticum

Common names:

Parrotfeather Brazilian watermilfoil

Origin: Amazon River basin

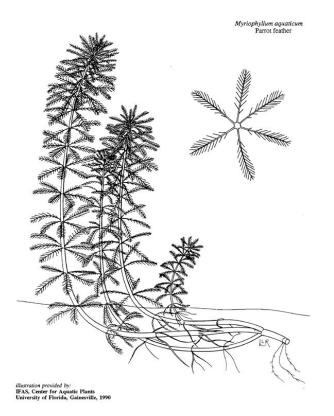
Key features:

Plants occur mostly above the water's surface **Stems:** Thick red stems, sometimes green **Leaves:** Leaves are a blue-green color and have a feathery appearance, leaves are whorled, dissected with rounded tips **Flowers:** Flowers have white sepals and no petals (only

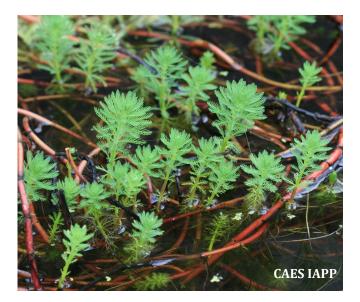
females found in the US) Fruits/Seeds: 0.06-0.08 inches (1.5-2 mm) long Reproduction: Fragmentation

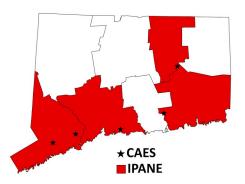
Easily confused species:

Eurasian watermilfoil: *Myriophyllum spicatum* Variable-leaf watermilfoil: *Myriophyllum heterophyllum*









Page 20 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Myriophyllum heterophyllum

Common names:

Variable-leaf watermilfoil Variable watermilfoil Two-leaf watermilfoil

Origin:

Southern United States

Key features:

Plants are submersed

Stems: Dark brown stems extend to the water's surface and spread to form large mats

Leaves: Triangular with ≤ 11 pairs of leaflets. Leaves are dissected and whorled (4-6 leaves/whorl) resulting in a feathery appearance with leaf whorls < 1 inch apart giving it a ropy appearance

Flowers: Inflorescence spike 2-14 inches (5-35 cm) long extend beyond the water's surface with flowers in whorls of four with reddish petals

Fruits/Seeds: Fruits are almost round, with a rough surface

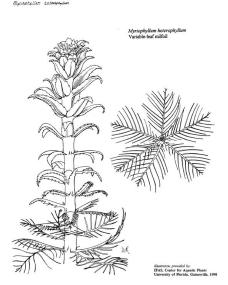
Reproduction: Fragmentation and seeds

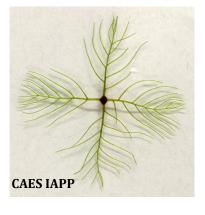
Easily confused species:

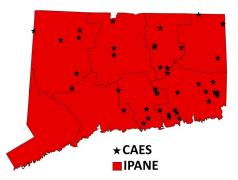
Eurasian watermilfoil: *Myriophyllum spicatum* Low watermilfoil: *Myriophyllum humile*











Myriophyllum spicatum

Common name:

Eurasian watermilfoil

Origin:

Europe and Asia

Key features:

Plants are submersed

Stems: Stem diameter below the inflorescence is greater with reddish stem tips

Leaves: Leaves are rectangular with ≥ 12 pairs of leaflets per leaf and are dissected giving a feathery appearance, arranged in a whorl, whorls are 1 inch (2.5 cm) apart

Flowers: Small pinkish male flowers that occur on reddish spikes, female flowers lack petals and sepals and have 4 lobed pistil

Fruits/Seeds: Fruit are round 0.08-0.12 inches (2-3 mm) and contain 4 seeds

Reproduction: Fragmentation and seeds

Easily confused species:

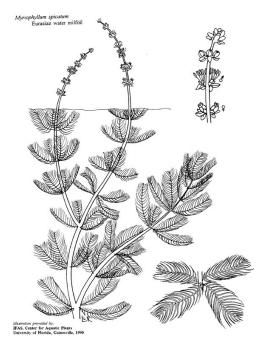
Variable-leaf watermilfoil: *Myriophyllum heterophyllum* Low watermilfoil: *Myriophyllum humile* Northern watermilfoil: *Myriophyllum sibiricum* Whorled watermilfoil: *Myriophyllum verticillatum*



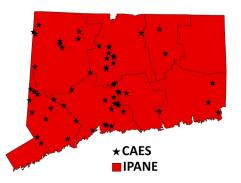








Copyright 1991 Univ. of Florida Center for Aquatic and Invasive Plants



Najas minor

Common names:

Minor naiad Brittle waternymph Spiny leaf naiad Eutrophic waternymph

Origin:

Europe

Key features:

Plants are submersed

Stems: Branched stems can grow up to 4-8 inches (10-20 cm) long

Leaves: Opposite and lance shaped on branched stems with easily visible toothed leaf edges and leaves appear curled under, basal lobes of leaf are also serrated, 0.01-0.02 inches (0.3-0.5 mm)

Flowers: Monoecious (male and female flowers on same plant)

Fruits/Seeds: Fruits are purple-tinged and seeds measure 0.03-0.06 inches (1.5-3 mm)

Reproduction: Seeds and fragmentation

Easily confused species:

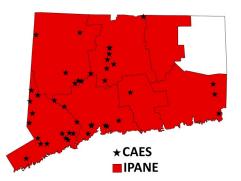
Other naiads (native): Najas spp.











Nelumbo lutea

Common name: American water lotus

Origin:

Southeastern United States, Mexico, Honduras, and the West Indies

Key features:

Plants are on or above the water

Stems: Stiff stalk attaches to the center of the leaf

Leaves: Large, bluish-green, circular leaves with no "slit" like water lilies

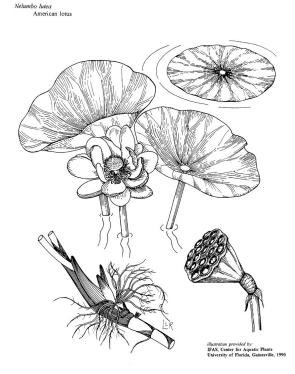
Flowers: White to yellowish flowers measure up to 8 inches (20 cm) wide

Fruits/Seeds: Seeds are nut-like and contained in a structure that resembles the top of a watering can

Reproduction: Seed

Easily confused species:

None



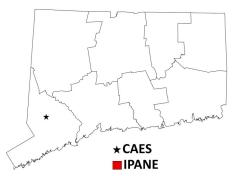
Copyright 1991 Univ. of Florida Center for Aquatic and Invasive Plants



Shaun Winterton, Aquarium and Pond Plants of the World, Edition 3, USDA APHIS PPQ, Bugwood.org







Nymphoides peltata

Common name: Yellow floating heart

Origin: Europe, Japan, China, and India

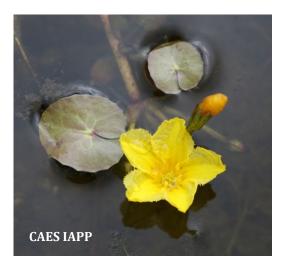
Key features:

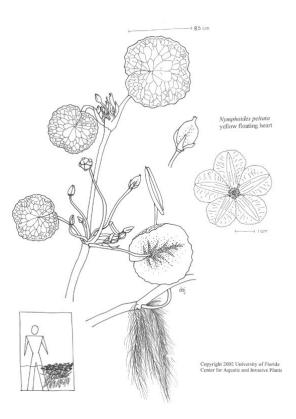
Floating leaf plant Stems: Branching stems spread over water's surface Leaves: Floating leaves are round and heart-shaped at base, paired at each node Flowers: Flowers are bright yellow on long peduncles with 5 fringed petals Fruits/Seeds: Seeds are flat and oval and are in capsules Reproduction: Seeds and rhizomes

Easily confused species:

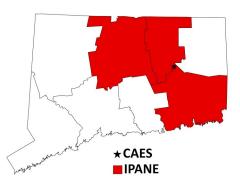
Little floating heart: *Nymphoides cordata* Yellow water lily: *Nuphar variegata*







Copyright 2002 Univ. of Florida Center for Aquatic and Invasive Plants



Pistia stratiotes

Common names:

Water lettuce Tropical duckweed

Origin:

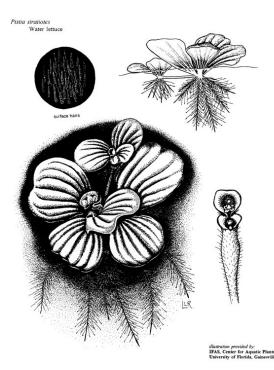
Nativity unknown, but possibly South America, Africa, Southeastern US

Key features:

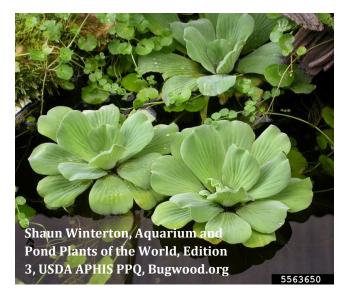
Free floating plant that resembles a head of lettuce
Stems: Roots are long and feathery
Leaves: Leaves are fleshy and covered with dense white hairs and have parallel venation
Flowers: Several male flowers form a whorl around a spike with one female flower below them
Fruits/Seeds: Fruit are light green berries that pro- duce 0.04 inch (1 mm) brown seeds
Reproduction: Seeds and stolons

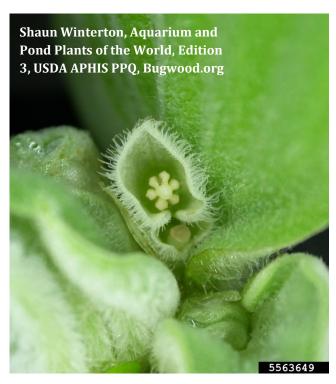
Easily confused with:

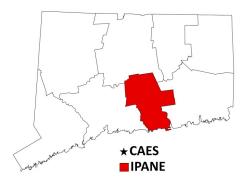
None



Copyright 1991 Univ. of Florida Center for Aquatic and Invasive Plants







Page 26 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Potamogeton crispus

Common names:

Curly leaf pondweed Crispy-leaved pondweed Crisped pondweed

Origin:

Asia, Africa, and Europe

Key features:

Plants are submersed

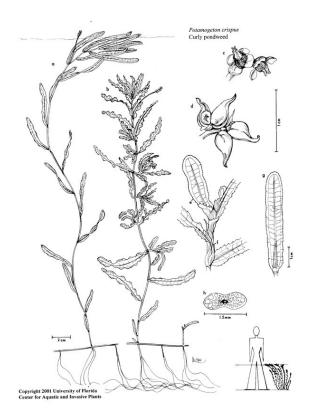
Stems: Stems are flattened, can form dense stands in water up to 15 feet (5 m) deep

Leaves: Alternate leaves 0.3-1 inches (3-8 cm) wide with wavy edges (similar to lasagna) with a prominent mid-vein

Flowers: Brown and inconspicuous Fruits/Seeds: Fruit is oval 0.1 inches (3 mm) long Reproduction: Turions (right) and seeds

Easily confused species:

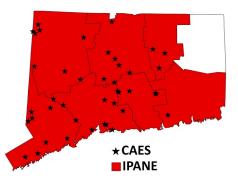
None











Rorippa microphylla

Synonym: Nasturtium microphyllum

Common name:

Onerow yellowcress

Origin:

North Africa, Europe, and the Middle East

Key features:

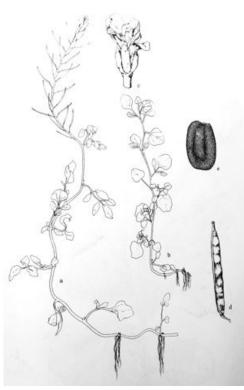
Stems: Grows flatly across the ground and roots at nodes, forming large mats, can be fully to partially submerged
Leaves: Pinnate leaves with 3-9 segments and the terminal leaf is the largest
Flowers: White petals with 4 part perianth

Fruits/Seeds: Fruit is a long and slender silique, up to 1 inch (25

mm) long, with seeds in one row on each side **Reproduction**: Seed

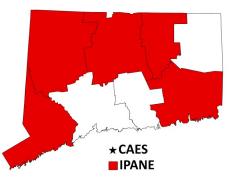
Easily confused species:

Watercress: Rorippa nasturtium-aquaticum



Crow and Hellquist, 2000







Page 28 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Rorippa nasturtiumaquaticum

Synonym:

Nasturtium officinale

Common name:

Watercress

Origin:

North Africa, Europe, and the Middle East

Key features:

Stems: Hollow stems can grow flat on mud or be fully or partially submersed

Leaves: Leaves are pinnately compound have 3-9 segments and vary in shape, the terminal leaf is the largest in each segment **Flowers:** Small white and green flowers; four white petals with four long and 2 short stamens

Fruits/Seeds: Fruit is pod-like silique, 0.4-0.6 in. (10-15 mm) long, with seeds in two rows per side

Reproduction: Fragmentation and seed

Easily confused species:

Onerow yellowcress: Rorippa microphylla





Connecticut, Bugwood.org







Salvinia molesta

Common names:

Giant salvinia Water fern Salvinia Kariba weed Aquarium watermoss

Origin:

Brazil

Key features:



Stems: Horizontal stems float below the surface **Leaves:** Submersed leaves are brown and feather-like; surface leaves are folded at midrib and covered with many water repellent hairs that are split in the middle but rejoin at the tips; leaves become tightly packed into long chains as the plant grows

Fruits/Seeds: Egg shaped sporocarps Reproduction: Fragmentation

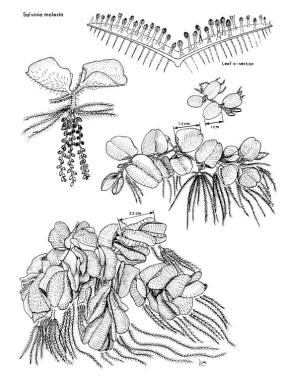
Free floating plant with no roots

Easily confused species:

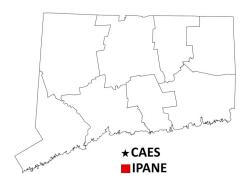
None



Photo Credit A: Mic Julien, Commonwealth Scientific and Industrial Research Org., Bugwood.org B: Vic Ramey Copyright 2000 University of Florida



Copyright 2002 Univ. of Florida Center for Aquatic and Invasive Plants



Trapa natans

Common names:

Water chestnut European water chestnut

Origin: Asia and Europe

Key features:

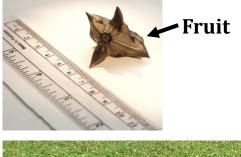
Plants are rooted to substrate and float **Stems:** Stem is submersed, flaccid and can be up to 15 feet (5 m) long **Leaves:** Leaves 0.8-0.16 inches (2-4 cm) long are triangular and toothed along the front edge with inflated petioles, leaves float in a rosette pattern **Flowers:** Flowers are located in the center of the rosette and have four white petals **Fruits/Seeds:** Fruit is hard and has four sharp

Fruits/Seeds: Fruit is hard and has four sharp spines

Reproduction: Seeds and fragmentation

Easily confused species:

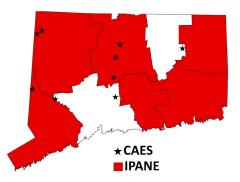
None





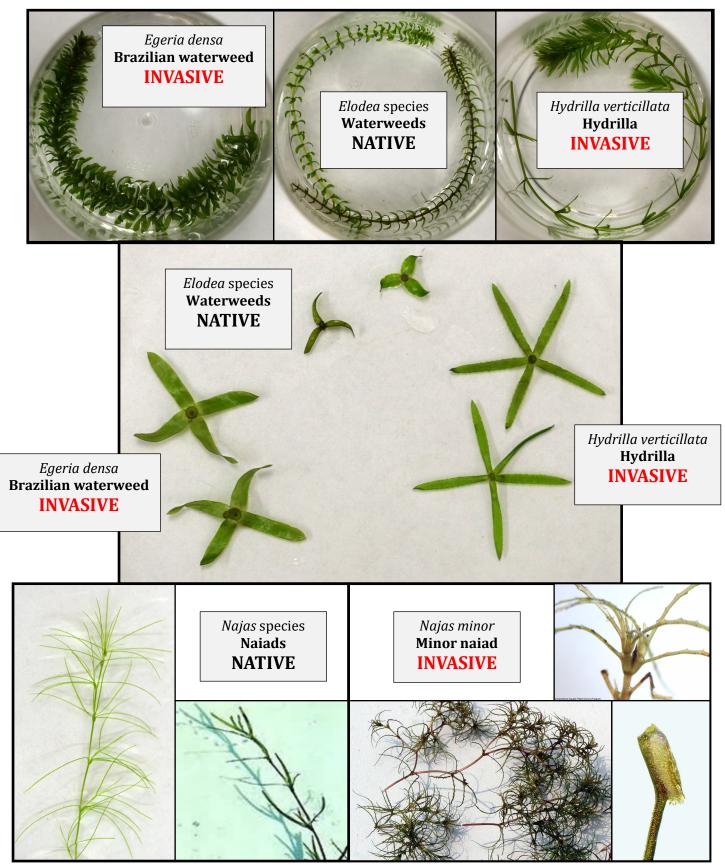






Commonly Confused Aquatic Plants Submersed plants with non-dissected leaves

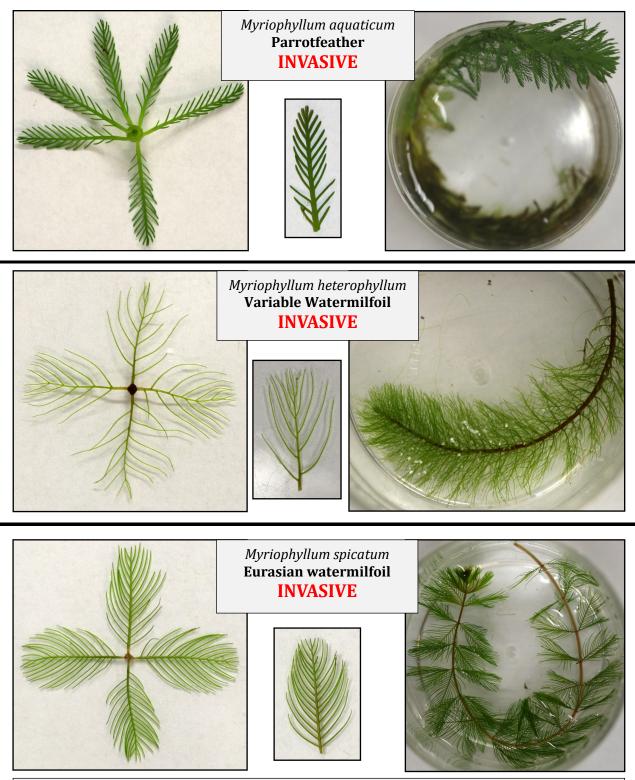
(all photos by CAES IAPP)



Page 32 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Submersed plants with feathery dissected leaves

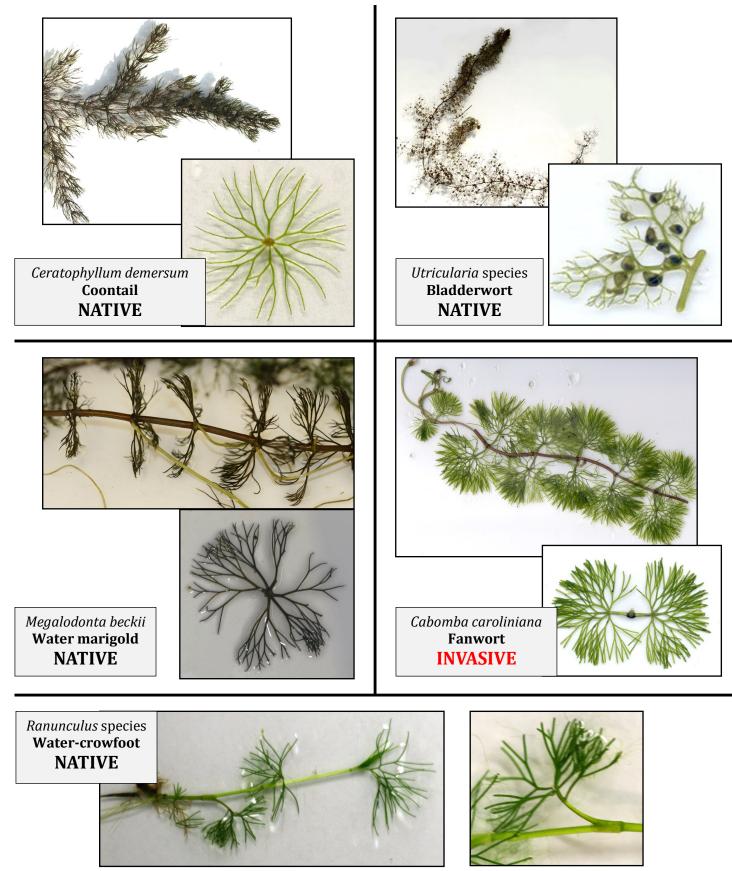
(all photos by CAES IAPP)



NOTE: *Myriophyllum sibiricum*, **Northern Watermilfoil**, is a threatened native species that is easily confused with *M. spicatum*. *M. sibiricum's* distinguishing features include less than 12 leaflet pairs per leaf, winter buds, and stem tips that are usually green instead of red.

Submersed plants with forked and branched dissected leaves

(all photos by CAES IAPP except where noted)



Page 34 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Key to Invasive or Potentially Invasive Aquatic Plants of Connecticut

Key also includes commonly confused native species

Floating-Leaf Plants (field characteristics)

1. Plants free-floating on water's surface, not rooted to the substrate*
2. Leaves folded along midrib, surface covered with hairs
2. Leaves not folded, surface smooth
3. Petioles inflated; oval leaves in a rosette; light purple flowers
Eichhornia crassipes (Water Hyacinth)
3. Petioles not inflated; broad, fleshy leaves in a rosette, covered with dense white hairs
1. Plants rooted in substrate
4. Leaves forming a rosette; leaves triangular, toothed; petioles inflated; spiny fruit
4. Leaves not forming rosette
5. Leaves compound, cut into several leaflets
6. Leaves comprised of four leaflets, like a four-leaf clover
Marsilea quadrifolia (European Waterclover)
6. Leaves pinnately compound with 3-9 leaflets, terminal leaflet is largest; hollow
stems floating; small white and green flowers in clusters
7. Pod-like fruit 0.4-0.6 inches (10-15 mm) long, 2 rows of seeds per side
Rorippa nasturtium-aquaticum (Watercress)
7. Pod-like fruit 0.7-1 inches (17-26 mm) long, 1 row of seeds per side
5. Leaves entire or lobed
8. Leaves entire (no slit), circular, bluish green, on stiff stalk above water
8. Leaves lobed, heart shaped
9. Yellow flowers
10. Flowers with five, fringed petals Nymphoides peltata (Yellow Floating Heart)
10. Flowers ball shaped, petiole flattened Nuphar variegata (Yellow Water Lily) (native)
9. White flowers with five, fringed petals; roots close to the floating leaves, near the surface of the
water

*Plants such as yellow and little floating heart and water chestnut can become free-floating when dislodged from sediment or detached from a rooted plant.

Submersed Plants (field characteristics)

 Leaves entire, sometimes toothed Leaves alternate, with wavy edges (lasagna-like); turions may be present; prominent leaf mid vein
<i>Potamogeton crispus</i> (Curly Leaf Pondweed)
2. Leaves whorled, opposite, or clustered
3. Leaf bases wider than the leaf blade, appearing opposite, whorled or clustered
4. Toothed leaf edges visible without magnification
4. Magnification needed to see toothed edge
3. Leaf base not distinct from rest of leaf blade, leaves strictly whorled
5. Whorls of 3 leaves; leaf margins not toothed <i>Elodea</i> species (Waterweeds) (native)
5. Whorls of 4 or more leaves; leaf margins toothed (magnification sometimes needed)
6. Leaves 4 per whorl (rarely up to 6 leaves/whorl), 0.5-1.5 inches (1.2-4 cm) long, toothed leaf
margins (need magnification) <i>Egeria densa</i> (Brazilian Waterweed)
6. Leaves 5 per whorl (rarely 2-6 leaves/whorl), 0.2-0.7 inches (0.6-1.7 cm) long, toothed leaf
margins; mid-vein may be toothed; tubers present; may have turions
7. Leaves feathery in appearance (pinnate)
8. Leaves concentrated above the water; thin, rounded-tipped, blue-green leaves
<i>Myriophyllum aquaticum</i> (Parrotfeather)
8. Leaves concentrated below the water, except for emergent flower spikes
9. Leaf whorls less than 1 inch (2.5 cm) apart, giving the plant a ropy look; triangular shaped leaves,
with less than or equal to 11 pairs of leaflets; thick spike with entire toothed leaves
Myriophyllum heterophyllum (Variable Watermilfoil)
9. Leaf whorls 1 inch (2.5 cm) apart; rectangular shaped leaves, with greater than or equal to 12
pairs of leaflets; thin spike with leaves smaller than flowers
<i>Myriophyllum spicatum</i> (Eurasian Watermilfoil)
9. Leaves rounded in whorls with less than 12 pairs of leaflets; winter buds
10. Leaves with numerous small bladders, not rooted <i>Utricularia</i> species (Bladderworts) (native)
10. Leaves lacking bladders
11. Leaves alternate; petioles sheathing stem; flowers usually solitary
11. Leaves opposite or whorled
12. Leaves whorled; leaf divisions fork in pairs, forking a total of 1-4 times, leaves often toothed;
no roots or flower spike Ceratophyllum species (Hornworts) (native)
12. Leaves opposite, fan-shaped; leaf divisions fork into either 2 or 3 segments
13. Leaves attached to the stem with petioles; small floating leaves; flowers white
13. Leaves not attached to the stem by a petiole, leaves opposite but appearing whorled;
emersed leaves on spike entire to toothed; flowers yellow

Introduction and Dispersal of Aquatic Invasives

(all photos by CAES IAPP)

Aquariums and Water Gardens

Most of the invasive aquatic plants in Connecticut are thought to have been introduced via the release from aquariums or water gardens (Les and Mehrhoff 1999). Although setting these plants "free" rather than disposing of them sounds like a good idea, it should be avoided. Because it is difficult to know what species you have and the designation of "non-invasive" is not always accurate, all aquarium and water gardening plants should be treated as invasive and disposed of properly. This includes, returning to a retailer, giving to well-informed aquarist or water gardener, or sealing in plastic bags for disposal in the trash. Also be aware that water gardens that are upstream from waterbodies are prone to releasing aquatic plants,



fish and other species downstream during flooding events.



Clean, Drain, Dry

Proper boat and trailer cleaning is an important way to prevent the spread of aquatic invasive species. Many plants spread through fragmentation, so even a small piece stuck to a trailer can create an infestation in a waterbody. Zebra mussels and other aquatic animals can also hitch a ride on boats and trailers to new locations. Best practices for boat and trailer cleaning include the Clean, Drain, Dry technique and disinfection. Clean all equipment, removing any visible mud, plants, fish or animals before leaving the launch site. Drain all water from boats and live

wells at the launch site. **Dry** anything that comes into contact with water for a minimum of five days or wash with hot water that is at least 140 degrees F for a minimum of 30 seconds (preferably at high pressure). Boats, trailers, and equipment can be sprayed with a bleach solution (3 oz. per gallon) allowing ten minutes of contact time prior to rinsing. Equipment can also be dipped into 100% vinegar for 20 minutes. The CT DEEP has more information on aquatic hitchhikers (<u>https://portal.ct.gov/DEEP/Fishing/General-Information/Aquatic-Invasive-Species</u>).

The "Clean, Drain, Dry" technique can prevent the spread of invasive species.

(Credit - https://portal.ct.gov/DEEP/Fishing/General-Information/Aquatic-Invasive-Species)



Clean:

- Inspect and remove all visible plant, fish, and animals as well as mud or other debris at the launch. Do not transport them home.
- Check trailer, including axle and wheel areas in and around the boat itself: anchor, props and jet engines, ropes, boat bumpers, paddles, and anything else that came in contact with the water.

Drain:

- Eliminate all water from every conceivable space and item before you leave the area you are visiting.
- Remove the drain plug from boats and put boat on an incline so that the water drains out
- Drain all water in live-wells, bilge, ballast tanks, transom wells, kayaks, canoes, rafts, motors, jet drives, boat hulls, scuba tanks and regulators, boots, waders, bait buckets, seaplane floats and swimming floats.

Dry:

• Dry Equipment, if possible, allow for 5 days of drying time before entering new waters.

Additionally, under the law

• Boaters must inspect their vessel for vegetation and aquatic nuisance species and properly remove and dispose of all vegetation and nuisance species before transporting the vessel. You can be fined \$95 per violation (CGA 15-180) and Public Act 12-167.

Managing Nuisance Aquatic Vegetation in Connecticut

(all photos by CAES IAPP)

Invasive Aquatic Plant Control

Aquatic vegetation proliferates in water with proper sunlight, chemistry, sediment and freedom from antagonistic organisms. Managing invasive aquatic plants usually requires a multifaceted approach. It is important to accurately identify the species present. This can be accomplished through an aquatic plant survey (top right). Sometimes nuisance plants, such as lily pads, are native. Although they may need to be managed, their removal could result in replacement with more problematic invasive species. In addition, certain native plants may be highly beneficial or rare and warrant protection. Combinations of management techniques that adapt to yearly changes are usually most effective. This discussion of invasive aquatic plant control is intended to be introductory in nature. The Aquatic Ecosystem Restoration Foundation (2020) has an online guidebook that details the subjects discussed below

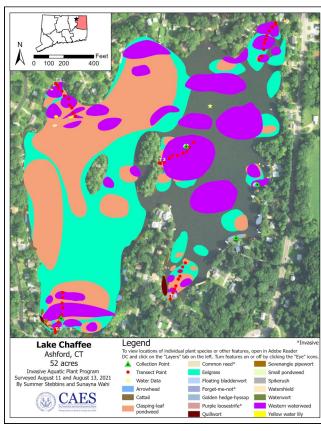
(http://aquatics.org/bmpchapters/BMP4ed.pdf).

Nutrient Reduction

Because nuisance aquatic vegetation is stimulated by nutrients, especially phosphorus and nitrogen, reducing the amount of these elements reaching a water body is an important part of any management program. Public education on preventing septic tank failures (bottom right) and use of fertilizers is im-

portant. Using soil tests to determine nutrient needs helps assure fertilizer is used at the proper rates and at the right time. Recent legislation in Connecticut has banned phosphorus from fertilizers used on established lawns unless substantiated by a soil test. CAES tests soil for citizens (portal.ct.gov/caes). Unfertilized vegetated buffer zones along shorelines are effective in limiting the movement of fertilizer to ponds and lakes. The misapplication of fertilizer to pavement is of concern because storm drains often discharge into lakes and ponds.





Water Level Drawdown

Lowering the water level of a lake or pond can expose unwanted vegetation to lethal drying and freezing conditions. Where water level drawdown is feasible, this is a cost effective aquatic plant management technique. Non- target plants and other aquatic organisms, however, can be negatively impacted. Usually drawdowns are performed in the winter when recreational use is minimal. Warm winters, snow cover and groundwater seepage can prevent necessary freezing and drying. Minor naiad (*Najas minor*) seems tolerant to drawdown probably because it re-



grows from seeds each year. Care needs to be taken to properly assess the refill time. If the body of water is not filled by spring problem vegetation can expand into areas where plant growth is normally limited by light penetration. A side benefit of winter drawdown is docks and other shoreline structures are protected. CAES IAPP has been monitoring the effects of the annual drawdowns on Candlewood Lake (above) since 2007 and found good control of Eurasian watermilfoil (*Myriophyllum spicatum*) in the drawdown year (CAES IAPP, 2018). Drawdowns may require permits from local, state or federal agencies.

Herbicides

Managing invasive aquatic plants with herbicides is sometimes necessary. An aquatic herbicide must meet strict requirements of the United States Environmental Protection Agency and then be registered in the state where it is used. In Connecticut, aquatic herbicides may not be applied without obtaining a permit from the Connecticut Department of Energy and Environmental Protection (CT DEEP). Choosing the best herbicide requires proper plant identification, a sense for the non-target species you want to protect, the time of year you want to treat, potential water use limitations (i.e. drinking, swimming, irri-



gation etc.), and acceptance by stakeholders. There are two general types of herbicides, contact and systemic. Contact herbicides are usually quick acting but have little effect on roots and reproductive propagules such as seeds, turions and tubers. Regrowth, therefore, can be expected. Systemic herbicides are slower acting but have the capability of controlling the root system. Longer term control is possible but elimination of an invasive species is unlikely. The suppressive effect of herbicides often prevents further spread. Herbicides are available in liquid or granular forms (above). CT DEEP (2014) offers an online guide (<u>https://portal.ct.gov/-/media/DEEP/pesticides/Certification/Supervisor/aweedspdf.pdf</u>) to the latest approved aquatic herbicides and their use.

Sediment Removal

Shallow areas of lakes and ponds with fertile sediment will promote plant growth. Removal of the sediment is a long term solution. Sediment removal is performed through various types of dredging and permits from local, State and Federal agencies are usually necessary. Dredging is performed either wet or dry. Wet dredging does not require lowering the water level and has the advantage of minimal disruption to recreational use. Dry dredging (right) involves lowering the water level, drying the sediment and using excavation equipment to facilitate removal. This method is efficient and sometimes the material can be sold to



recoup some of the costs. The downside of dry dredging is its negative effects on the aquatic ecosystem and the inability to use the waterbody for long periods (often many years).

Biological Controls

Introducing an organism that feeds specifically on an invasive aquatic plant can provide targeted longterm control. Unfortunately, virtually no reliable target specific biocontrols are available for invasive aquatic plants in the northeastern USA. A biocontrol being tested for Eurasian watermilfoil is the milfoil weevil (*Euhrychiopsis lecontei*) (bottom). Although the larvae and adults feed on Eurasian watermilfoil and

they are native to most lakes with the plant, neither the natural or introduced populations are currently considered a reliable control. The most common biocontrol in Connecticut is an herbivorous fish called grass carp (*Ctenopharyngodon idella*) (right). This fish was originally cultivated in China for food, but its propensity to eat vegetation brought it to Europe and the USA for aquatic weed control. Concerns over grass carp developing breeding populations have caused it to become regulated in many states including Connecticut. Prior to liberation, a state permit must be obtained, the fish must be documented to be sterile (triploid) and inlets and outlets usually must be screened to prevent the fish





from escaping. Grass carp

are usually considered more suitable for smaller waterbodies and are introduced at various rates depending on the amount of vegetated acres, the plants being controlled and other factors. Recently, however, these fish have been used in larger lakes including Candlewood Lake, Connecticut's largest lake. The fish take a year or two to obtain sufficient size to reduce vegetation and after about five years restocking is often necessary. Unfortunately grass carp may prefer native plant species and unforeseen damage to the aquatic ecosystem can occur. In addition, as plants are consumed and passed through the fish's digestive system nutrients are released into the water that may cause algal blooms. Because grass carp are sensitive to copper based algaecides, treating the algae can be challenging.



Harvesting

Probably the simplest means for controlling an area of invasive aquatic plants is removing them by hand pulling or mechanical cutting (above, left). Hand pulling is particularly effective in small areas and can reap tremendous benefits when used to remove new infestations. Sometimes SCUBA divers are employed. Unfortunately, many areas are too large for hand pulling to be practical and mechanical cutters (above, right), rakes or suction harvesters are needed. If the root systems are not removed rapid regrowth can occur, and the procedure may be considered an expensive mistake. Several lakes in Connecticut have dedicated weed harvesting boats that operate each year (above, right). To prevent new introductions, weed harvesters need to be thoroughly cleaned before moving from one waterbody to another.

Benthic Barriers

Benthic barriers are blanket-like materials that are spread over unwanted aquatic vegetation to prevent light from reaching the plants. They are particularly well suited to small areas but occasionally are used for larger areas. Although most benthic barriers are installed in the spring and removed in the fall, they can be installed for as little as several weeks and then moved. Work done by CAES IAPP has shown that benthic barriers can be effective when installed for as little as a month. More research is needed to document the level of control when this procedure is utilized. If benthic barriers are left in place for more than one growing



season sediment settles on the surface and plants can take root.

Sources of further information on invasive aquatic plant management

Aquatic Ecosystem Restoration Foundation. 2020. Biology and Control of Aquatic Plants – A Best Management Practices Handbook Fourth Edition. L. Gettys, W. Haller, D. Petty, eds. Retrieved May 11, 2022 from <u>http://aquatics.org/bmp.html</u>

CT DEEP. 2014. Nuisance Aquatic Vegetation Management - A Guidebook. Retrieved May 11, 2022 from https://portal.ct.gov/-/media/DEEP/pesticides/Certification/Supervisor/aweedspdf.pdf

Nuisance & Purportedly Invasive Algae in Connecticut Waterways

Algae are important members of aquatic ecosystems. They can range from uni-cellular micro-algae, diatoms, (cannot see with the naked eye) to 300 foot tall kelp forests of macro-algae. They are photo-autotrophs, meaning they fix CO₂ through photosynthesis in order to make energy. As a result of photosynthesis, they release oxygen. Many diatoms, including the 3 species that will be discussed here, are indicator species of water quality and overall environmental conditions because of how sensitive they can be to things like temperature, pH, and available nutrients. For this reason, algae and diatoms have been reported to be at great risk to climate change and recently, algae have been reported in areas not previously thought to be. Invasive diatom species are causing damage to eco-systems and infrastructure in Connecticut waterways and abroad.



Cymbella janischii

Cymbella janischii was first reported in the Farmington River in 2011. It is a known invasive species in Connecticut and can be a nuisance because of its ability to form thick matts of growth on the bottom of rivers and streams. It is unsightly, can cause a disruption to recreational activities such as swimming, and disrupts the benthic ecosystems by covering large amounts of the area it grows in. *Cymbella janischii* is native to the pacific northwest of the United States but is not indigenous to Connecticut. It has been reported in areas as far away as Japan. Compared to the *Didymosphenia* taxa, *Cymbella janischii* prefers warmer waters

with higher nutrients. It is hypothesized that *Cymbella janischii* spreads on fishing gear, specifically felt waders that provide a dark, moist area for the algae to survive in until the next use in

a different waterbody. Moving this algae can be avoided by rinsing fishing gear with water that's over 140 degrees Fahrenheit for at least 30 seconds or spraying with a solution of bleach and water (3 oz / gallon) for at least 10 minutes. Mucilaginous stalks of *Cymbella janischii* are pictured to the right covering a rock.



Didymosphenia geminata—Rock Snot, Didymo

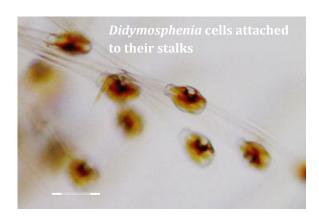
First reported east of the Mississippi in 2005, *Didymosphenia geminata* easily covers 100% of benthic ecosystems in rivers and streams with thick matts of algal growth. Also known as rock snot, this algae is present in the Farmington River where fishermen noticed it growing on rocks. Didymo is known to thrive in waterways that have low nutrient levels, specifically low phosphorus. It can thrive in man made waterways designed for things like hydro power



plants, water treatment plants, canal systems, and agricultural waterways for irrigation. In addition to this, didymo is extremely unsightly and is a nuisance where it blooms. Because of its destructive nature to benthic ecosystems and its impact on important infrastructure, control measures have been investigated. As with *Cymbella janischii*, care should be taken to clean fishing gear after using it in order to stop it spreading into other waterbodies.

Didymosphenia hullii— Rock Snot, Didymo

Didymosphenia hullii was reported as a new species in 2016 and looks very similar to *D. geminata*. It requires identification under a microscope to tell the two apart. *Didymosphenia hullii* and other species are genetically similar, and *D. hullii* is perhaps evolutionarily young.





Content for this section including all images was provided by Dr. Diba Khan-Bureau. Citations can be found in the Literature Cited section of this document.



Didymosphenia hullii mucilaginous stalks



Invasive Aquatic Clams and Mussels

Invasive freshwater Asian clams (*Corbicula fluminea*) and zebra mussels (*Dreissena polymorpha*)are now present in many Connecticut waterbodies. Quagga mussels (*Dreissena bugensis*) have been found in neighboring states and will probably spread to Connecticut. Like invasive aquatic plants, these bivalves can colonize rapidly and degrade natural ecosystems, fisheries, recreational opportunities, and real estate values. In addition, these organisms can threaten mechanisms used for hydroelectric power generation. Zebra mussels colonize hard surfaces including boats and docks, and soft surfaces such as aquatic plants (see figure below). Species identification for these three bivalves can be found on the following pages.

There are a variety of native mussel and clam species that are abundant in Connecticut lakes and ponds. Generally, the native species are larger than the ones described in this guide. Refer to the CTDEEP guide to native mussels for further information (<u>https://portal.ct.gov/-/media/DEEP/wildlife/pdf files/nongame/fwmuslpdf.pdf</u>).



Corbicula fluminea

Common names:

Golden clam Asian clam Asiatic clam Good luck clam

Origin:

Asian Africa Australia

Key features:

At the sediment surface or slightly buried Size: 0.5 to 0.75 inches Shell: triangular shape (unlike most oval-shaped clams), beak is centrally located and high Color: Yellowish brown to black Reproduction: Hermaphroditic, self-fertilization

Easily confused species:

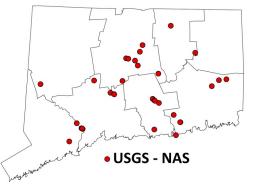
Fingernail clams: Sphaeriidae





5530924





Page 46 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Dreissena bugensis

Common name: Quagga mussel

Origin: Dnieper River drainage of Ukraine

Key features:

Colonizes on hard surfaces Size: up to 1.5 inches Shell: unstable on edge (unlike *Dreissena polymorpha*), round shape, asymmetrical Color: dark concentric rings, color is paler near the hinge , black, cream, or white bands



Amy Benson, U.S. Geological Survey, Bugwood.org UGA1265039

Easily confused species:

Zebra mussel: Dreissena polymorpha





Rebekah D. Wallace, University of Georgia, Bugwood.org 5489098

• USGS - NAS

Dreissena polymorpha

Common name: Zebra mussel

Origin:

Black, Caspian, and Azov Seas (Europe)

Key features:

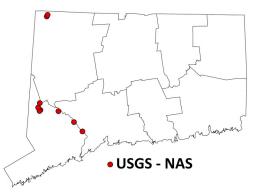
Colonizes on hard surfaces Size: less than one inch Shell: conspicuous black stripes, stable on flattened underside (unlike *Dreissena bugensis*) Color: color patterns vary, dark or light colored shells, most commonly with stripes

Easily confused species:

Quagga mussel: Dreissena bugensis





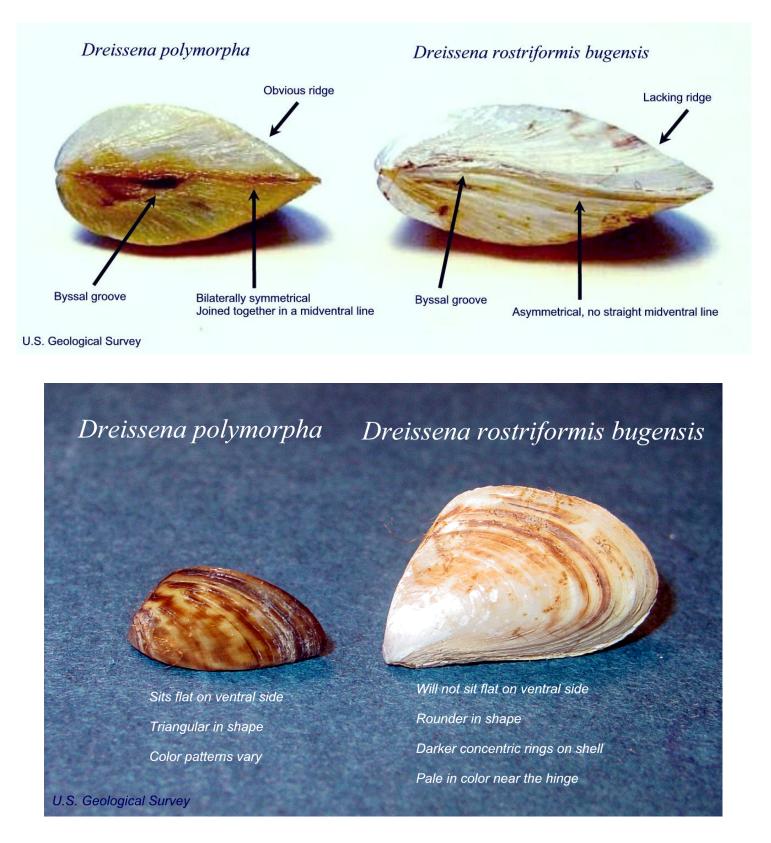




Page 48 - Connecticut's Invasive Aquatic Plant, Clam, and Mussel Identification Guide

Commonly Confused Aquatic Mussels Differences between Quagga and Zebra Mussels

(all photos by Myriah Richerson—USGS-NAS)



Literature Cited

Aquatic Ecosystem Restoration Foundation. 2020. Biology and Control of Aquatic Plants – A Best Management Practices Handbook Fourth Edition. L. Gettys, W. Haller, D. Petty, eds. Retrieved May 11, 2022 from http://aquatics.org/bmp.html

Aquatic Plant Management Website. 2013. Aquatic plant fact sheet. Retrieved March 1, 2018. http://www.weedscience.ncsu.edu/aquaticweeds/.

Britton, N.L. and A. Brown. 1913. An illustrated flora of the northern United States, Canada and the British Possessions. Vol. 1:37.

CAES IAPP. 2018. The Connecticut Agricultural Experiment Station Invasive Aquatic Plant Program (CAES IAPP). Retrieved March 1, 2018. http://www.ct.gov/caes/iapp.

Center for Aquatic and Invasive Plants, University of Florida. 2005. Aquatic, Wetland and Invasive Plants in Pen-and-Ink. IFAS Publications, Gainesville, Florida.

Center for Aquatic and Invasive Plants, University of Florida. 2018. Plant information and images. Retrieved March 1, 2018. http://plants.ifas.ufl.edu/

Connecticut Aquatic Nuisance Species Working Group. 2006. Connecticut aquatic nuisance species management plan. Retrieved March 3, 2018. http://www.ctiwr.uconn.edu/ProjANS/SubmittedMaterial2005/ Material200601/ANS%20Plan%20Final%20Draft121905.pdf.

Connecticut Department of Energy and Environmental Protection. 2018. Aquatic Invasive Species. Retrieved March 5, 2018 http://www.ct.gov/deep/cwp/view.asp?a=2696&q=322690&deepNav_GID =1630&pp=12&n=1.

Connecticut Department of Energy and Environmental Protection. 2013. A Field Guide to the Freshwater Mussels of Connecticut. Retrieved March 5, 2018. http://www.ct.gov/deep/lib/deep/wildlife/ pdf_files/nongame/fwmusl.pdf.

Cornell University. 2018. New York Invasive Species Information. Retrieved March 3, 2018. http://nyis.info.

Couch, R. and E. Nelson. 1985. Myriophyllum spicatum in North America. Pp. 8-18 in: Anderson, L.W.J., ed., Proceedings of the First International Symposium on Water Milfoil (Myriophyllum spicatum) and related Haloragaceae Species. Aquatic Plant Management Society, Vicksburg, Mississippi.

Crow, G.E. and C.B. Hellquist. 2000. Aquatic and wetland plants of northeastern North America. Vol 1. Pteridophytes, gymnosperms, and angiosperms: dicotyledons. The University of Wisconsin Press, Madison, Wisconsin.

Crow, G.E. and C.B. Hellquist. 2000. Aquatic and wetland plants of northeastern North America. Vol 2. Angiosperms: Monocotyledons. The University of Wisconsin Press, Madison, Wisconsin.

Fassett, N. 1957. A manual of aquatic plants. Rev. Eugene Ogden. The University of Wisconsin Press. Madison, Wisconsin.

Fishman, K.J., R.L. Leonard and F.A. Shah. 1998. Economic evaluation of Connecticut lakes with alternative water quality levels. Connecticut Department of Environmental Protection. 79 Elm St. Hartford CT.

Global Invasive Species Database. 2018. Species information. Retrieved March 3, 2018. http://www.icungisd.org/.

Invasive and aquatic species. 2018. Weeds. Invasive.org. Retrieved March 1, 2018. http://www.invasive.org/.

Invasive plant atlas of New England (IPANE). 2018. IPANE species. Retrieved March 1, 2018. http://www.eddmaps.org/Species/

Invasive plant atlas of New England. 2018. Data and Maps. Retrieved March 1, 2018. https://www.eddmaps.org/ipane/.

Khan-Bureau, D. A., Blanco, S., Morales, E. A., Ector, L., Beauchene, M. S., & Lewis, L. A. 2022. Using geometric morphometrics to disentangle Didymosphenia hullii and D. geminata (Bacillariophyceae) from Connecticut, USA, and two congeneric relatives. European Journal of Phycology, 1-11.

Khan-Bureau, D. A., Morales, E. A., Ector, L., Beauchene, M. S., & Lewis, L. A. 2016. Characterization of a new species in the genus Didymosphenia and of Cymbella janischii (Bacillariophyta) from Connecticut, USA. European Journal of Phycology, 51(2), 203-216.

Khan-Bureau, D. A., Beauchene, M. S., Ector, L., Morales, E. A., & Lewis, L. A. 2014. Observations of two nuisance stalk-forming diatoms (Bacillariophyta) from a river in Connecticut, Northeastern USA: Didymosphenia sp. and Cymbella janischii (A. Schmidt) De Toni. BioInvasions Record, 3(3).

Les, D.H., and L.J. Mehrhoff. 1999. Introduction of nonindigenous aquatic vascular plants in southern New England: A historical perspective. Biological Invasions 1:281-300.North Carolina State University

Northeast Aquatic Nuisance Species Panel. 2018. Species information. Retrieved March 3, 2018. http://northeastans.org.

Tenaglia, D. Missouri Plants. 2007. Retrieved March 3, 2018. http://www.missouriplants.com/.

Texas Invasive Species Institute. 2014. Species database. Retrieved March 3, 2018. http://www.tsusinvasives.org.

United Water Conservation District. 2010. Species information. Retrieved March, 3 2018. http://www.unitedwater.org.

USDA National Invasive Species Information Center. 2018. Aquatic Species. Retrieved March 3, 2018. https://www.invasivespeciesinfo.gov/aquatics/main.shtml.

USDA Natural Resources Conservation Service (NRCS). 2018. Plants database. March 3, 2018. http://plants.usda.gov/.

USGS Nonindigenous Aquatic Species. 2018. Species information. Retrieved March 3, 2018. http://nas.er.usgs.gov.

Washington State Noxious Weed Control Board. 2010. Weed list. Retrieved March 1, 2018. http://www.nwcb.wa.gov/.

Notes

Notes

Notes

The Connecticut Agricultural Experiment Station was founded in 1875. It is chartered by the General Assembly to make scientific inquiries and conduct experiments regarding plants and their pests, insects, soil and water, and to perform analyses for state agencies. Station laboratories or research farms are located in New Haven, Windsor, Hamden, and Griswold.



Equal employment opportunity means employment of people without consideration of age, ancestry, color, criminal record (in state employment and licensing), gender identity or expression, genetic information, intellectual disability, learning disability, marital status, mental disability (past or present), national origin, physical disability (including blindness), race, religious creed, retaliation for previously opposed discrimination or coercion, sex (pregnancy or sexual harassment), sexual orientation, veteran status, and workplace hazards to reproductive systems unless the provisions of sec. 46a-80(b) or 46a-81(b) of the Connecticut General Statutes are controlling or there are bona fide occupational qualifications excluding persons in one of the above protected classes. To file a complaint of discrimination, contact Dr. Jason White, Director, The Connecticut Agricultural Experiment Station, 123 Huntington Street, New Haven, CT 06511, (203) 974-8440 (voice), or Jason.White@ct.gov (e-mail). CAES is an affirmative action/equal opportunity provider and employer. Persons with disabilities who require alternate means of communication of program information should contact the Chief of Services, Michael Last at (203) 974-8442 (voice), (203) 974-8502 (FAX), or Michael.Last@ct.gov (e-mail).